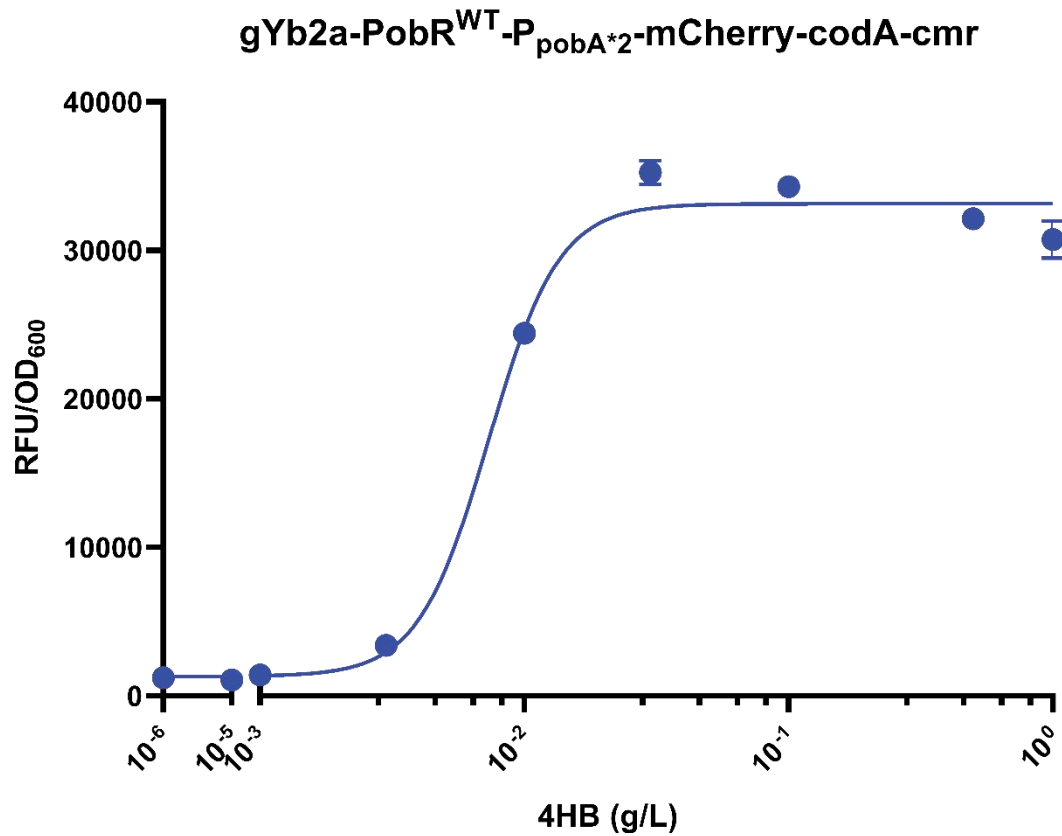


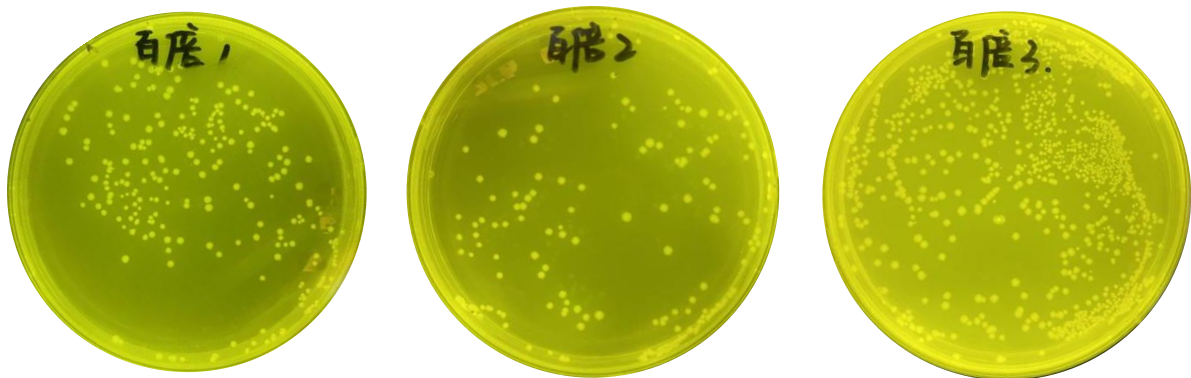
Supplementary Materials

Supplementary Figure 1



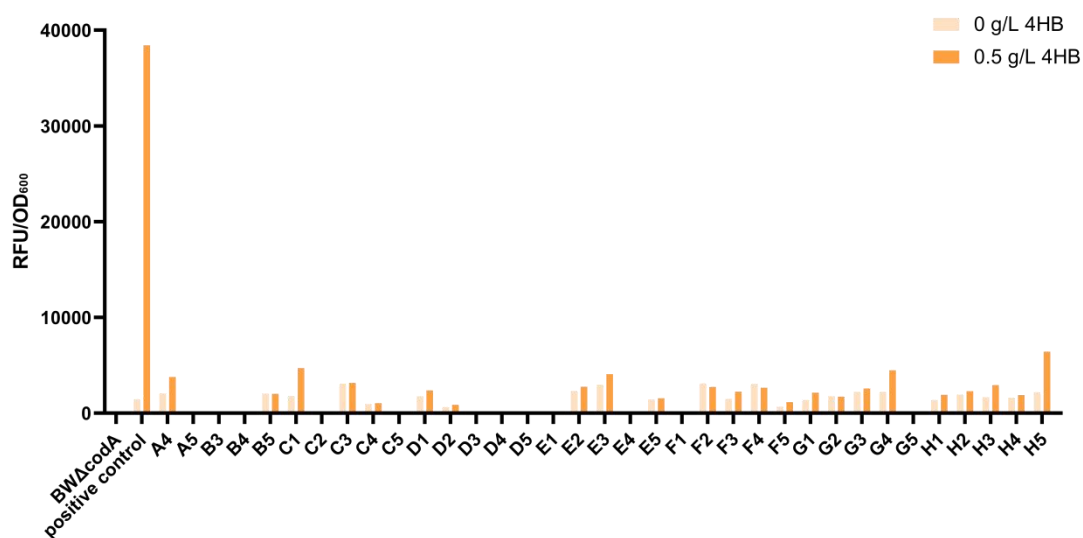
Supplementary Figure 1. The dynamic range of the PobR^{WT} Biosensor (in *E. coli* BWΔ*codA*) responsive to 4HB. The vertical axis is the ratio of the reporter gene *mCherry* (RFU) expression to the growth of *E. coli* (OD₆₀₀), measured after a 12 h of cultivation in M9 medium. Each value represents the mean ± standard deviation from 3 biological replicates.

Supplementary Figure 2



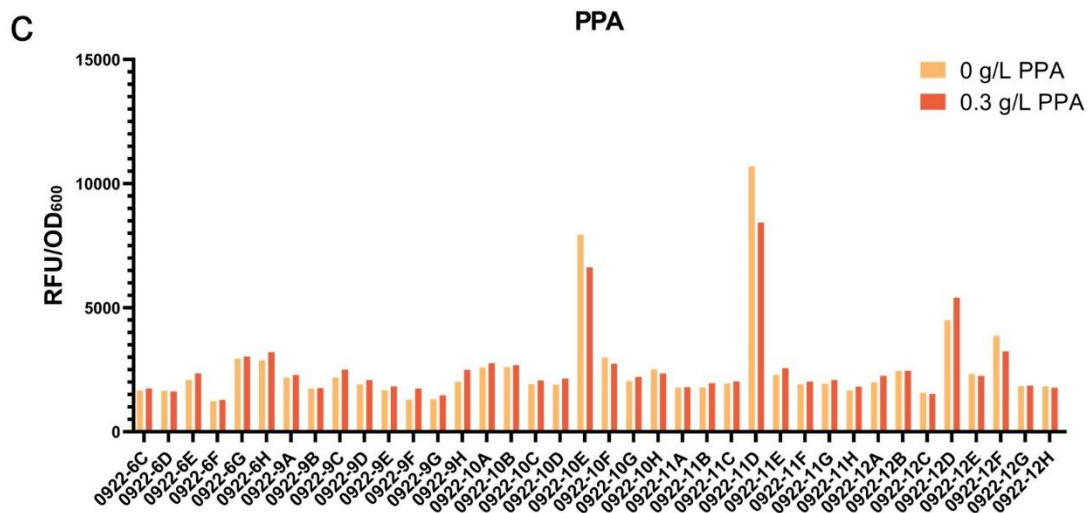
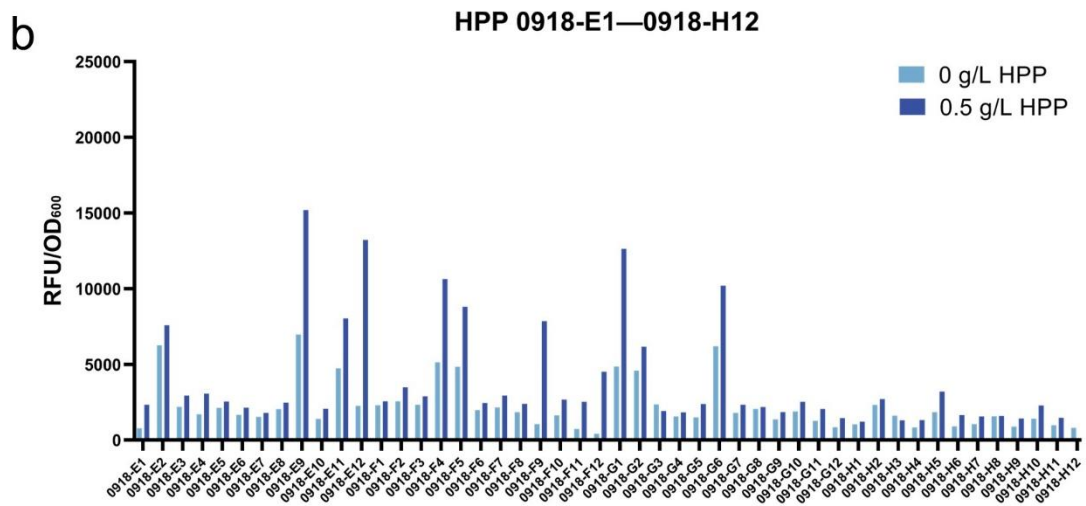
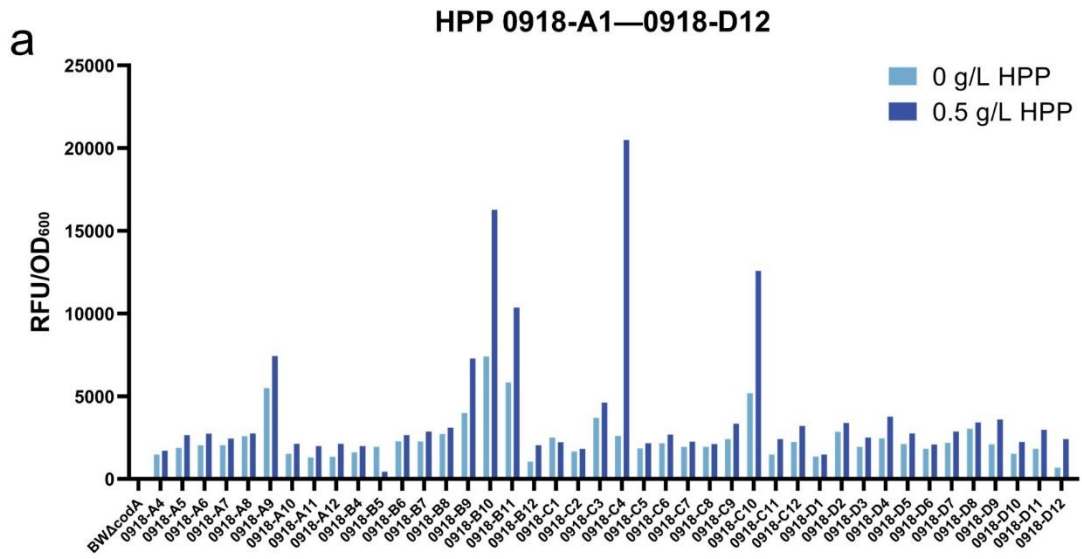
Supplementary Figure 2. Clone calculation after passing two rounds of negative selection in medium containing 50 mg/L and 200 mg/L 5-FC using the dilution coating method. The experimental procedure includes aspirating 50 μ l of the selected bacterial solution, adding it into 5000 μ l of LB medium (i.e., 1:100 dilution), and then taking 50 μ l of the diluted bacteria to spread onto a plate. Based on our calculation, the original density of the two-round selected bacteria was 450,000 CFU/mL.

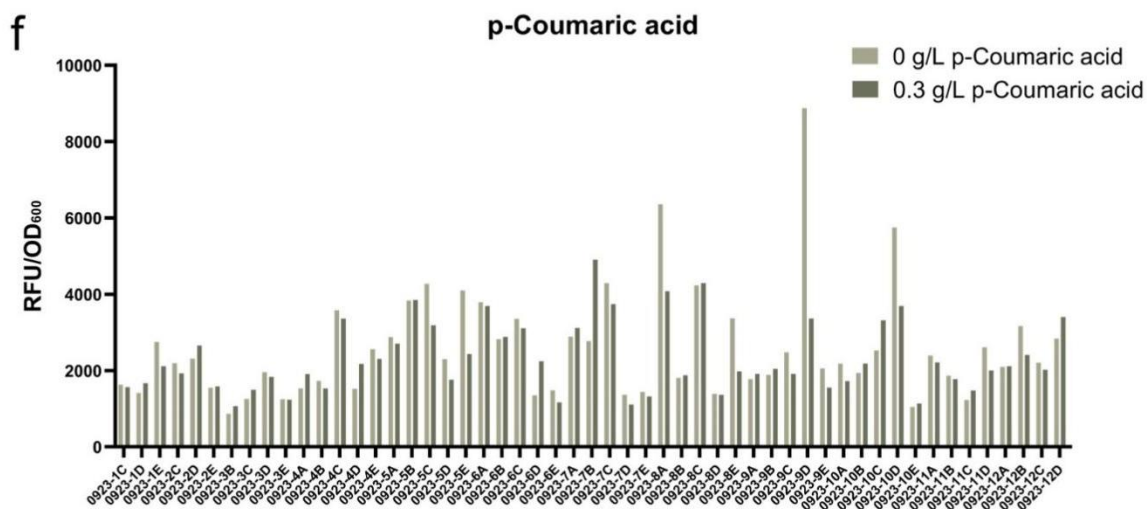
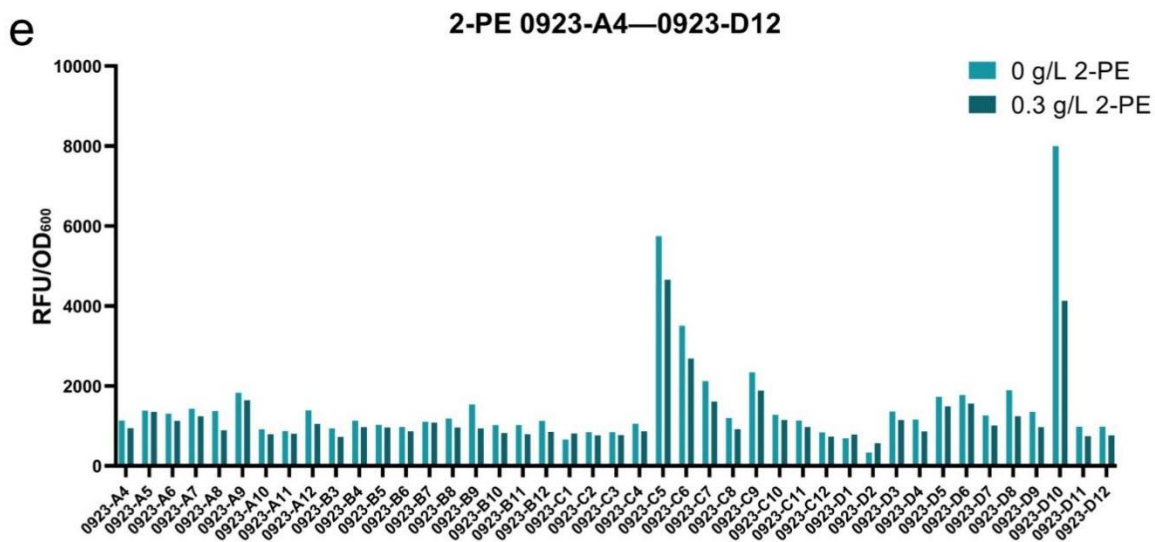
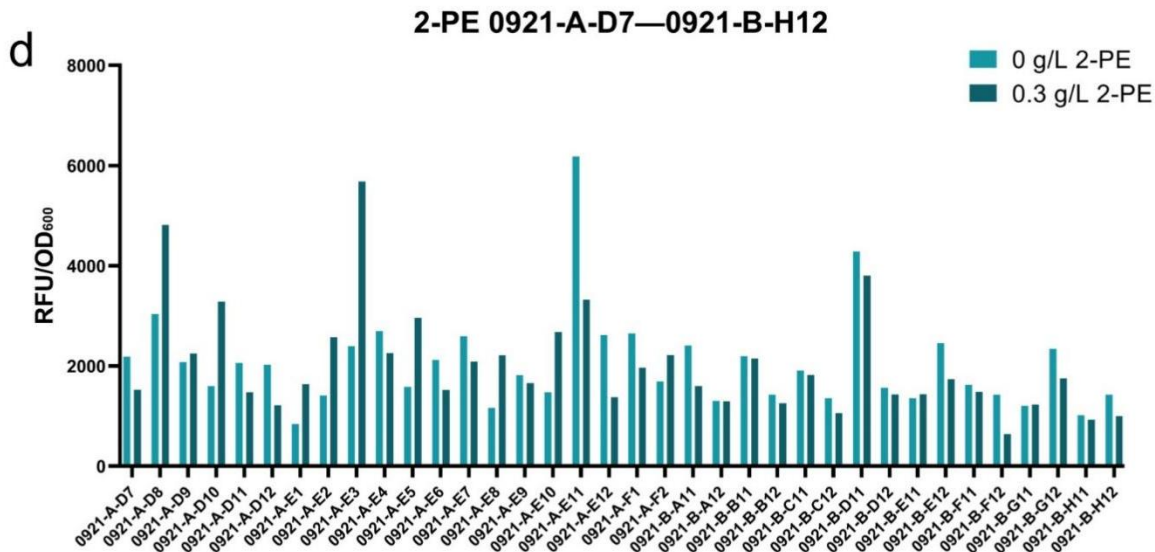
Supplementary Figure 3



Supplementary Figure 3. Response of the PobR strains to 4HB after two rounds of negative selection (50 mg/L and 200 mg/L). Thirty-five single colonies were randomly picked after diluted bacterial culture was spread on LB plates containing only ampicillin. They were activated by LB medium for 10 h and then transferred to M9 medium containing 0.5 g/L 4HB for 12 h of cultivation.

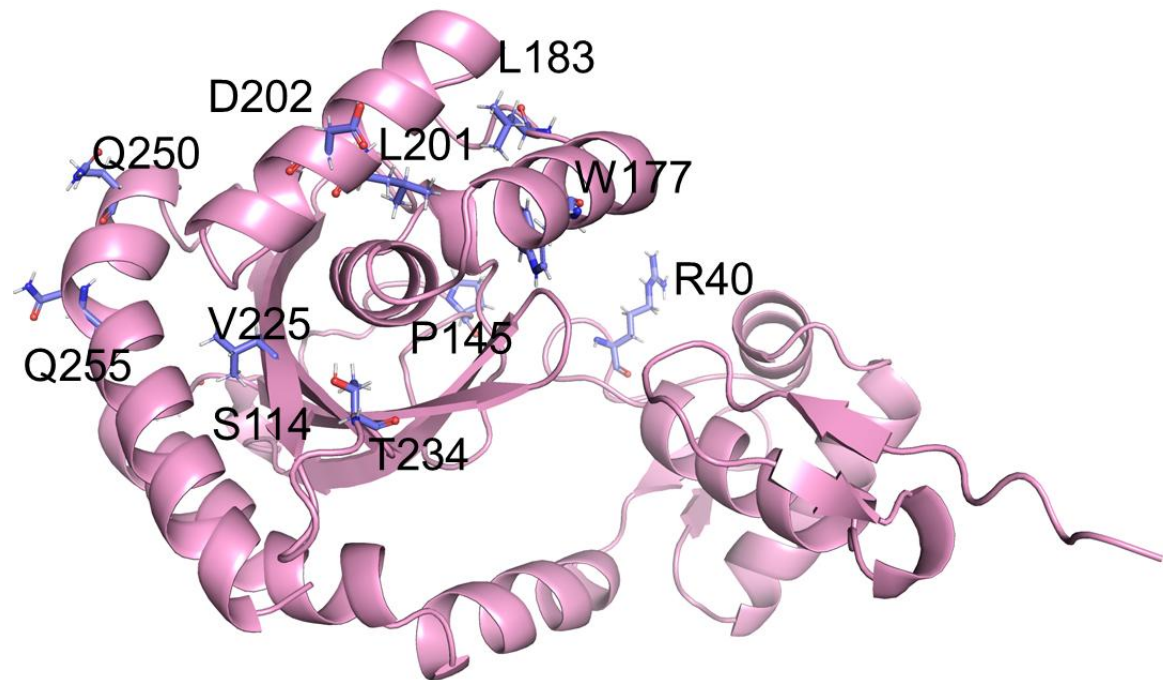
Supplementary Figure 4.





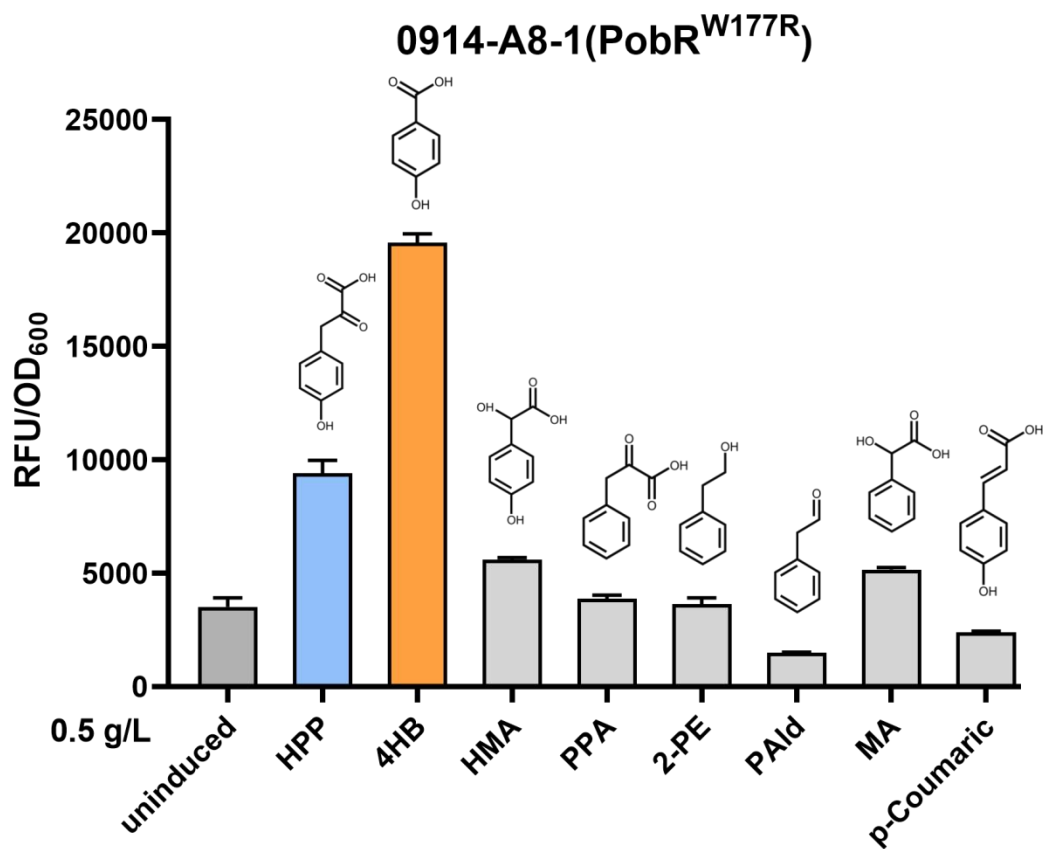
Supplementary Figure 4. Preliminary selection results. (a) to (i) Evaluation of individual clones for their responsiveness to HPP (a and b), PPA (c), 2-PE (d and e), p-Coumaric acid (f), PAld (g), MA (h), and HMA (i). The colonies or clones are denoted by their screening/selection identification numbers. The mean fold induction in specific mCherry fluorescence in response to the presence of various ligands serves as a measure to compare the PobR mutant biosensors. The concentration of the compounds was 0.3 g/L, except for HPP and PPA. The mean fold induction in specific mCherry fluorescence in response to the presence of various ligands serves as a measure to compare the PobR mutant biosensors. The concentration of the compounds was 0.3 g/L, except for HPP and PPA.

Supplementary Figure 5.



Supplementary Figure 5. Analysis of the position of amino acid mutation sites using the PyMOL.

Supplementary Figure 6.



Supplementary Figure 6. Fluorescence changes of the 0914-A8-1 clone (i.e., PobR^{W177R}) to the treatments of different aromatic compounds with structural similarity to HPP.

Supplementary Table 1. Bacterial strains and plasmids used in this study.

Strain	Description	Source
DB3.1	<i>F- gyrA462 endA1 glnV44</i> <i>Δ(sr1-recA) mcrB mrr hsdS20(rB-, mB-) ara14 galK2 lacY1 proA2 rps L20 (SmR) xyl5 Δleumt1</i>	Lab stock
BW25113Δ <i>codA</i>	<i>rrnBT14 ΔlacZWJ16 hsdR514</i> <i>ΔaraBAD_{AH33} ΔrhaBAD_{LD78}ΔcodA</i>	Lab stock
plasmid	Description	Source
gYB2a-ccdb	<i>Amp^r, P15A ori, ccdb gene</i>	Lab stock
pYB1a-eGFP-cmr	<i>Amp^r, P15A ori, eGFP gene, Chl^r</i>	Lab stock
pUAM-RE-CD	<i>Amp^r, Anderson J23100-promoter, RE gene, codA gene</i>	Lab stock
pYP1a- P _{pobA} *2-mCherry-sacB	<i>Amp^r, P15A ori, two P_{pobA} promotor, pobR gene, mCherry gene, sacB gene</i>	Lab stock
gYB2a-P _{pobA} *2-mCherry-sacB	<i>Amp^r, P15A ori, two P_{pobA} promoters, mCherry gene, sacB gene</i>	this study
gYb2a-P _{pobA} *2-mCherry-SacB-cmr	<i>Amp^r, P15A ori, two P_{pobA} promotor, mCherry gene, sacB gene, Chl^r</i>	this study
gYb2a-P _{pobA} *2-mCherry-CD-cmr	<i>Amp^r, P15A ori, two P_{pobA} promotor, mCherry gene, codA gene, Chl^r</i>	this study

gYb2a-PobR-mCherry-CD-cmr

Amp^r, P15A ori, two P_{pobA} promoter,
pobR gene, *mCherry* gene, *codA*
gene, *Chl^r*

this
study

Supplementary Table 2. Primers used in this study.

Primers	Sequences (5'-3')
PpobA*2-mc-0311-F	gagggctctcatccgagacgggtaccATTGGTGATGCTGTTCCAT
Primer 2-0311-R	ggatctcctgctgatgtcggaattcttattgtaactgtaatt
cmr-gibson-0317-F	gatctcctgctgatgtcggaattcttacgccccgccctgccact
cmr-gibson-0317-R	aattaacagttaacaaataagaattcatggagaaaaaatcactgg
CD-gibson-0420-R	ggtatatccagtgatttttctccatgaattctaccgttgtaatcgatgg
CD-gibson-0420-F	gtaactcgagaggagatgtcgaataacgctttac
cmr-Gibson-F	caaacggtaagaattcatggagaaaaaatcactggatatacc
Mc-Gibson-R	gacatctcctcgcgagtactgtacagctcgtccatg
PobR-P1-Bsal-F	gtgctgGGTCTCggATGGAACAGCATCACCAATA
PobR-P2-Bsal-R	agcgtgGGTCTCTGCTAAACCAAGTTGCGCAGTTCAT
